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# Single Ag atom engineered 3D-MnO<sub>2</sub> porous hollow microspheres for rapid photothermocatalytic inactivation of *E. coli* under solar light



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#### ABSTRACT

Atomic level Ag loaded MnO<sub>2</sub> porous hollow microspheres (Ag/MnO<sub>2</sub> PHMs) were prepared by redox precipitation method, and utilized for *E. coli* inactivation under solar light irradiation. Ag nanoparticles (NPs) can be downsized into single atoms, thereby realizing highly utilization of Ag element as well as achieving superior photothermocatalytic *E. coli* inactivation for Ag/MnO<sub>2</sub> than MnO<sub>2</sub> PHMs. Under attack by the optimal 0.3%Ag/MnO<sub>2</sub> PHMs with atomic Ag, 7.11 log<sub>10</sub> cfu/mL cells can be completely inactivated within 10 min, much faster than the 0.3%Ag/MnO<sub>2</sub> PHMs with Ag cluster (3.3 log<sub>10</sub> cfu/mL) prepared by photodeposition method, demonstrating the feasibility of redox precipitation to prepare efficient catalyst for water disinfection. Three effects are believed to contribute to this bacterial inactivation enhancement: (1) atomic Ag with high conductivity induces more formation of Mn<sup>3+</sup> and oxygen vacancies in MnO<sub>2</sub>, which can efficiently accelerate the separation of hot electrons and holes generated by MnO<sub>2</sub>, collectively work with itself generated hot electrons to form into reactive species for photocatalysis; (2) atomic Ag exhibits strong local heating effect and induces higher reducibility for MnO<sub>2</sub>, considerably enhances the photothermal conversion and lattice oxygen activity of MnO<sub>2</sub>, thus promoting the thermocatalysis; and (3) the synergism of solar light driven photocatalysis and thermocatalysis through the activated O<sub>L</sub>. The highly efficient photothermocatalysis make the designed 3D atomic Ag/MnO<sub>2</sub> PHMs have a promising antibacterial ability for cleaning the microbial contaminated water environment.

## 1. Introduction

Worldwide, more than half of billion people are reliant on a drinking water source contaminated with pathogenic microorganisms, such as bacteria, fungi and viruses [1]. As a result, over half a million deaths each year related to the consumption of unsafe water, with a majority of deaths caused by waterborne illnesses like cholera, typhoid, and diarrhea [2,3]. Chemical disinfection methods like chlorination and ozonation are considered as the most practical solution to provide safe drinking water, but they need to consume specific materials, and present risks to the promotion of persistent antibiotic resistance and formation of disinfection byproducts [4–6]. In contrast, photocatalytic disinfection has been considered as a sustainable and cost-effective alternative process, because of its lower cost, high efficiency, recyclable, limited disinfection byproducts, and the capability of full use of solar

energy [7-10].

Advanced photocatalysts that can effectively harness solar energy for water treatment have been extensively explored, involving modified  ${\rm TiO_2}$  and  ${\rm non\text{-}TiO_2}$  based materials, such as metal oxides, sulfides, and  ${\rm non\text{-}metal}$  g-C<sub>3</sub>N<sub>4</sub>, red phosphorus, *etc.* [11–14]. As a n-type semiconductor, earth abundant MnO<sub>2</sub> showed widely application for pollutants elimination with attractive features such as low-cost, chemical inertness, photo-stability and nontoxicity [15]. Among these properties, its highly efficient solar driven thermocatalytic ability through the transformation between Mn<sup>4+</sup> and Mn<sup>3+</sup> and promotion of oxygen vacancies is possibly the most important [16]. In general, catalysts with different morphologies or structures exhibit different physical and chemical properties [17]. For example, MnO<sub>2</sub> has been synthesized into three-dimensional (3D) nanostructures such as rods, tubes, spheres and sheets, which obtained great applications in catalytic fields [18–20].

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Recently, considerable attentions have been paid to the developing mesoporous materials on account of their large specific surface area, interconnected channels and abundant active sites [21]. Especially, 3D  $\alpha$ -MnO<sub>2</sub> porous hollow microspheres (MnO<sub>2</sub> PHMs) and its modified nanocomposites have been extensively studied for catalytic ozonation [22] and anti-fouling membrane [23].

The metal oxides doped with plasmonic noble metal (Ag, Au and Pt, etc.) could achieve remarkably enhanced catalytic ability, such as Au/ CeO<sub>2</sub>, Au/MnO<sub>2</sub> [24,25]. The composite exhibited obvious advantages because they combine the optoelectronic properties of semiconductor with the surface plasmon resonance (SPR) effect and excellent conductivity of noble metal [26]. The SPR effect could increase its light absorption ability by scattering resonant photons and the conductivity could accelerate the separation of charge carriers [27,28]. Most importantly, the plasmonic nanometal can provide driving forces to catalytic reactions by generating hot electrons due to its photothermal effect, which will beneficial for the photocatalysis reaction [29,30]. Some past studies have explored the use of SPR photothermal metals (e.g., Au, Ag, Cu and Al) to thermally inactivate pathogens, including photodynamic therapy and biofilm control applications [31,32]. Therefore, integrating photothermal effect into photocatalysis should be promising for utilizing the solar energy to achieve highly efficient disinfection and its mechanism has not been identified well till now [33].

Recently, single-atom catalysis has become a hot research filed, because the promoted dispersion of noble metal via downsizing a nanoparticle to the atomic level can maximum enhance its utilization [34,35]. However, the modification of catalysts with atomic noblemetal remains a tough task, because the noble-metal nanoparticles with high surface energy easily agglomerate into large particles through conventional co-precipitation or photo-deposition [36-38]. Inspired by the strategy for stabilizing single-atom Au on the surface of MnO<sub>2</sub> rods via defect trapping by Chen et al. [39], a modified redox precipitation was developed to fabricate single-atom Ag-deposited 3D a-MnO<sub>2</sub> PHMs in the present study. With this preparation method, the as-prepared Ag/ MnO<sub>2</sub> PHMs with the amount of Ag loading reduced to only 0.3% can efficiently use full solar spectrum, visible-infrared, and up to infrared light for E. coli inactivation. The broad-spectrum solar light utilization by Ag nanoparticle is related to the highly dispersed single-atom Ag as evidenced by Cs-corrected HAADF-STEM and EDS mapping. The synergistic inactivation by photocatalysis and thermocatalysis was substantiated and would be more desirable than photocatalysis alone for engineering practice.

# 2. Experimental

#### 2.1. Materials

All chemicals used in this work are analytical grade and used without any further purification. Silver nitrate (AgNO<sub>3</sub>) and Mn (CH<sub>3</sub>COO)<sub>2</sub>·4H<sub>2</sub>O were purchased from Tianjing Yongda Chemical Reagent Co. Ltd, Na<sub>2</sub>CO<sub>3</sub> were purchased from Shantou Xilong Chemical factory, KMnO<sub>4</sub> were purchased from Tianjing Fuchen Chemical Reagent Co. Ltd., and HCl (37%) were purchased from Guangzhou Reagent Chemical factory, respectively. In addition, the doubly-distilled deionized water was used throughout this study.

### 2.2. Preparation of Ag/MnO<sub>2</sub> porous hollow microspheres (PHMs)

 $MnO_2$  PHMs:  $MnCO_3$  precursor was prepared by mixing the Mn  $(CH_3COO)_2$ · $4H_2O$  and  $Na_2CO_3$  solutions, then  $MnO_2$  microspheres were formed. After that, HCl (10 mM) was added to remove the MnCO<sub>3</sub> core, and then calcined at 400 °C for 4 h to obtain 3D MnO<sub>2</sub> PHMs [22,23].

 $Ag/MnO_2$  PHMs: The preparation step was described in Scheme 1.  $MnO_2$  PHMs was ultrasonically re-dispersed in deionized water, and then  $AgNO_3$  solution was added. The concentrated  $H_2O_2$  (0.5 g, 30 wt.

%) was dropwise added to the mixture of MnO2 PHMs and AgNO3. During the addition of  $H_2O_2$ ,  $O_2$  was emitted.  $MnO_2 + H_2O_2 + 2H^+ =$  $Mn^{2+} + 2H_2O + O_2$ , where the consumption of released H<sup>+</sup> can drive the hydrolysis of AgNO<sub>3</sub> to Ag (OH) as equation:  $AgNO_3 + H_2O = Ag$ (OH) + HNO<sub>3</sub>. Because these two reactions occur simultaneously, the Ag species could be trapped by an in situ generated hole without aggregation. Moreover, Ag (OH) could be further reduced into metallic Ag by  $H_2O_2$ :  $O_2 + 2H^+ + 2e^- \rightleftharpoons H_2O_2$  and Ag (OH)  $+ H^+ + e^- \rightleftharpoons Ag + 2e^-$ H<sub>2</sub>O. After the redox precipitation, the solid was filtered, washed, and frozen-dried. Other Ag/MnO2 PHMs with different deposited amount were synthesized and denoted as (x) Ag/MnO<sub>2</sub>, x represents Ag/MnO<sub>2</sub> with different silver contents in preparation, which were 0.1, 0.3, and 0.5 wt. %, respectively. The accurate deposited Ag amounts for the asprepared samples were quantified by ICP-MS. Moreover, the PD-Ag/ MnO<sub>2</sub> PHMs prepared by photo-deposition method was also mentioned in the Text S1.

#### 2.3. Characterizations

The X-ray diffraction (XRD) patterns of samples were recorded on a Rigaku D/max 2200 PC diffractometer with Cu Kα radiation. UV-vis-NIR adsorption curve of catalysts were obtained using a UV-vis-NIR spectrophotometer (Lambda 950) with an integral-sphere attachment range from 200 to 1100 nm. XPS spectra of samples were collected on ESCALAB 250, Thermo Fisher Scientific, and USA. The morphology of samples was investigated by Quanta 400 F thermal field emission scanning electron microscopy (SEM, Shimazhu, Japan) using an acceleration voltage of 15 kV, and TEM (JEM-2010HR). The specific surface area for samples was carried out by an ASAP2010 physical and chemical adsorption instrument (USA). In addition, the photoelectrochemical performance was measured in a three electrode quartz cells system including a saturated calomel electrode (SCE) as reference electrode, platinum plate as counter electrode, and stainless steel coated with MnO<sub>2</sub> and Ag/MnO<sub>2</sub> PHMs as the working electrode with 1 M Na<sub>2</sub>SO<sub>4</sub> as electrolyte.

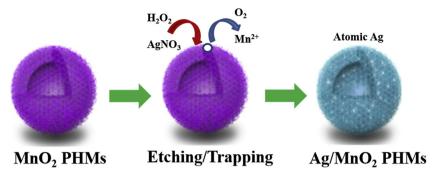
# $2.4. \ Photothermocatalytic, photocatalytic \ and \ thermocatalytic \ disinfection$

The bacteria were cultured in nutrient broth (NB) at  $37\,^{\circ}\mathrm{C}$  for  $16\,\mathrm{h}$  to yield a cell density of  $10^9\,\mathrm{cfu/mL}$ . Before test, the bacterial cells were washed twice by ultrapure water and then centrifugation ( $10,000\,\mathrm{rpm}$  for  $1\,\mathrm{min}$ ), and then diluted to  $10^7\,\mathrm{cfu/mL}$ . During each test,  $10\,\mathrm{mg}$  of catalyst and  $0.1\,\mathrm{mL}$  of bacteria solution ( $10^7\,\mathrm{cfu/mL}$ ) were added into a  $50\,\mathrm{mL}$  beaker, then the light source was turned on (a  $300\,\mathrm{W}$  Xenon). During each time interval, the collected sample was quickly spread on the agar plate and counted after  $24\,\mathrm{h}$  incubation at  $37\,^{\circ}\mathrm{C}$ .

Moreover, a long-wave pass cutoff filter of 420, 480, 560, 690, or 830 nm was used individually to analyze the photothermocatalytic activity of the Ag/MnO<sub>2</sub> PHMs under the visible-infrared or infrared irradiation from the Xenon lamp. The light intensity of the irradiation with  $\lambda > 420$ , 480, 560, 690, and 830 nm is 291.9, 273.3, 255.0, and 238.7, 206.2 mW/cm², respectively. To identify the photocatalytic activity of Ag/MnO<sub>2</sub> PHMs for *E. coli* inactivation, the reactor was placed in a water bath to maintain the ambient temperature under the irradiation of the Xenon lamp to eliminate the effect of thermal catalysis. To measure the thermocatalytic activity of the Ag/MnO<sub>2</sub> PHMs for *E. coli* inactivation, the reactor was placed in a water bath to maintain the temperature in the dark to eliminate the effect of photocatalysis.

#### 2.5. Microscopic observations of bacteria

(1) SEM: after interacting with particles for  $0\,\mathrm{min}$ ,  $60\,\mathrm{min}$  and  $120\,\mathrm{min}$ , the cells were collected by centrifuged at  $5000\,\mathrm{rpm}$  for  $5\,\mathrm{min}$ , then  $5\,\mathrm{mL}$  glutaraldehyde (2.5%) was added into the mixture to fix the cells overnight and washed twice with phosphate buffer saline (PBS) buffer, and then sequentially dehydrated with 30%, 50%, 70% and 90%



Scheme 1. Synthetic route of atomic Ag-doped MnO<sub>2</sub> PHMs.

ethanol for 10 min, and 100% ethanol for 20 min twice, respectively. Then specimen were lyophilized, gold sputter-coated to observe any changes in their morphologies on SEM; (2) Fluorescent-based cell live/dead test: all the bacterial samples (10<sup>7</sup> cfu/mL cells, 0.01 g/L catalysts) were stained with PI (propidium iodide) and SYTO9 of the Live/Dead Baclight Bacterial Viability kit, and then observed with a laser scanning fluorescence microscopy (Olympus, FV1000) [40].

# 2.6. Enzyme activity & Biomolecule oxidation assay

(1) The ATP synthesis ability of the treated cells at 5, 10, 20 and 30 min was monitored with the ATP assay kit, quantified by measuring absorbance at 630 nm; (2) Glutathione (GSH) was extracted using GSH-PX Assay Kit (Colorimetric method), then quantified by measuring absorbance at 630 nm. Samples were collected at 5, 10, 20 and 30 min; (3) Chromosomal DNA was extracted using an Ezup Column Bacteria Genomic DNA Purification Kit, then verified by DNA agarose gel electrophoresis (0.6% agarose gel at  $100\,\mathrm{V}$  for  $40\,\mathrm{min}$  in  $1\times\mathrm{TAE}$  buffer) [41].

#### 2.7. Photothermal effect measurement

To examine the photothermal effect induced by light irradiation,  $25\,\mathrm{mL}$  of  $1\,\mathrm{mg/mL}$  MnO $_2$  or Ag/MnO $_2$  PHMs dispersions were irradiated for  $30\,\mathrm{min}$  by a Xenon lamp or NIR laser ( $0.5\,\mathrm{W}$  cm $^{-2}$ ), respectively. The temperature changes of the solutions were monitored using a submerged thermocouple microprobe. Thermo images were also taken by a thermos imager (Testo Co., Ltd., Testo 885) to perform quantitative analyses of photothermal effect of samples. A testo IR soft was used to obtain the average temperature distribution for every thermos image. The parameters of NIR laser (Laserwave, Beijing, LWIRL808-5W-F) include wavelength:  $808\,\mathrm{nm}$ ; power:  $5\,\mathrm{W}$ ; power density  $0.5\,\mathrm{W/cm}^2$ ; the spot area is  $50\,\mathrm{mm}^2$  and the diameter is  $8\,\mathrm{mm}$ .

#### 3. Results and discussion

#### 3.1. Structural analysis of the catalysts

In this strategy (Scheme 1),  $\rm H_2O_2$  etches the surface of  $\alpha$ -MnO<sub>2</sub> PHMs through consuming released H<sup>+</sup>, thereby driving hydrolysis of AgNO<sub>3</sub> into Ag (OH), which could be then reduced into atomic Ag and subsequently trapped into surface defects of  $\alpha$ -MnO<sub>2</sub> to form atomic Ag/MnO<sub>2</sub> PHMs. First, the wide-angle XRD patterns of as-prepared catalysts were displayed in Fig. 1a. For all samples, the distinct diffraction peaks at  $2\theta = 12.7^{\circ}$ ,  $18.1^{\circ}$ ,  $28.8^{\circ}$ ,  $37.5^{\circ}$ ,  $42.1^{\circ}$ ,  $49.9^{\circ}$ ,  $56.2^{\circ}$ , and  $60.3^{\circ}$  can be well indexed to the tetragonal  $\alpha$ -MnO<sub>2</sub> phase (JCPDS 44-0141). In contrast to pure MnO<sub>2</sub> PHMs, the peak intensity of Ag/MnO<sub>2</sub> PHMs become slightly weaker but no additional peaks belonged to the crystal Ag can be found, presumably owing to the ultrafine-sized and high dispersion of nano-sized Ag [42]. Moreover, the BET surface areas decrease from 124.13 m<sup>2</sup> g<sup>-1</sup> for MnO<sub>2</sub> PHMs to 104.11 m<sup>2</sup> g<sup>-1</sup> for

0.3%Ag/MnO<sub>2</sub> PHMs (Table 1), confirming that Ag nanoparticles are highly dispersed and even partially filled into the porous MnO<sub>2</sub> PHMs. The high specific surface area and mesoporous structure of Ag/MnO<sub>2</sub> PHMs were conducive to the absorption and diffusion of reactants, which has a positive correlation with the amount of surface active sites [22,23]. Through the ICP-MS measurement, the loading amount of Ag in the 0.3%Ag/MnO<sub>2</sub> PHMs sample was determined to be 0.257 wt. % (Table 2).

The high-resolution XPS test was used to investigate the surface composition and chemical state of elements in the as-prepared catalysts. As shown in Fig. 1b, the Mn  $2p_{3/2}$  region was resolved into three individual sub-bands, represents Mn2+ (640.6 eV), Mn3+ (641.6 eV) and Mn<sup>4+</sup> (642.8 eV), respectively [43]. Table 2 indicates the surface concentration of  $Mn^{2+} + Mn^{3+}$  in 0.3%Ag/MnO<sub>2</sub> (1.06) is higher than that of MnO<sub>2</sub> (0.96), suggesting Ag doping can reduce Mn<sup>4+</sup> in MnO<sub>2</sub>. In general, the increase of low valent Mn can induce the formation of crystalline defects and oxygen vacancies in MnO2, which are beneficial for the photocatalysis/thermocatalysis through capturing more electrons and activating reactive species [44]. Moreover, all the O 1 s XPS spectra in Fig. 1c have three peaks at 529.7, 531.1 and 532.1 eV, assigned to lattice oxygen (O<sub>1</sub>), adsorbed oxygen (O<sub>ads</sub>, such as O<sub>2</sub><sup>-</sup>, O<sup>-</sup> and OH group) and limited surface oxygen (H<sub>2</sub>O), respectively [45]. Table 2 displays that molar ratio of O<sub>L</sub>: O<sub>ads</sub> in 0.3% Ag/MnO<sub>2</sub> PHMs (2.12) is higher than that of MnO<sub>2</sub> (1.78), evidencing the Ag doping can increase the formation of O<sub>L</sub>. Generally, O<sub>L</sub> with high mobility is favorable for the formation of reactive oxygen species through the transformation by oxygen vacancy [46]. Yang et al. also indicated that O<sub>L</sub> can be activated by solar light irradiation, which plays a crucial role in the thermocatalytic oxidation [47]. Additionally, there are two peaks belonging to Ag-3d $_{5/2}$  (368 eV) and Ag-3d $_{3/2}$  (374 eV) orbital, which can be further separated and fitted into Ag+ and Ag° species, respectively (Fig. 1d). This confirms the successful doping of Ag nanoparticles on MnO2 PHMs.

To inspect the defined structure of 0.3%Ag/MnO<sub>2</sub> porous hollow microspheres (PHMs) prepared by redox-precipitation method, SEM, HRTEM, HAADF-STEM, and EDS-mapping were utilized. The SEM images in Fig. 2a show that the 0.3%Ag/MnO<sub>2</sub> has regular 3D porous spherical shape, and the cavity of the broken MnO2 confirms its hollow structure. Similarly, the TEM in Fig. 2b also confirms the 3D hollow structure of 0.3%Ag/MnO<sub>2</sub> PHMs. As displayed in Fig. 2c, the HRTEM analysis of the 0.3%Ag/MnO2 PHMs reveals that the polycrystalline walls of MnO2 are grown along the [110], [310] and [211] directions, but it is hard to find Ag nanoparticles within catalyst. Interestingly, due to the difference in element contrast between Ag and Mn, the Ag species look brighter than the Mn in the dark field of Cs corrected HAADF-STEM (inset of Fig. 2c). It is found that Ag atoms (marked by white squares) obviously appearing at the edge of the MnO2 PHMs, evidencing that the method of redox precipitation can downsize Ag nanoparticles into single atoms. The HAADF-STEM image of 0.3%Ag/MnO<sub>2</sub> PHMs at larger scale in Fig. 2d shows no aggregation of the Ag species. The mapping images of Ag, Mn, and O shown in Fig. 2e-h further

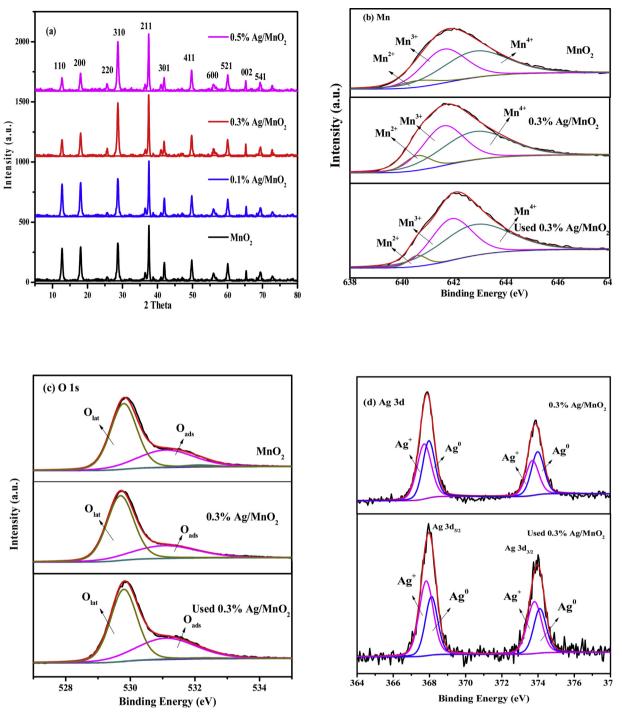


Fig. 1. (a) XRD patterns, XPS spectra of (b) Mn 2p, (c) O 1s, and (d) Ag 3d in  $MnO_2$  and  $Ag/MnO_2$  PHMs.

Table 1
Ag amount and specific surface area of MnO<sub>2</sub> and Ag/MnO<sub>2</sub> PHMs.

| Catalysts               | Ag content<br>wt.% | $S_{BET} m^2 g^{-1}$ |
|-------------------------|--------------------|----------------------|
| $MnO_2$                 | 0                  | 124.13               |
| 0.1%Ag/MnO <sub>2</sub> | 0.104              | 120.87               |
| 0.3%Ag/MnO <sub>2</sub> | 0.257              | 104.11               |
| 0.5%Ag/MnO <sub>2</sub> | 0.492              | 102.95               |

indicate that atomic Ag nanoparticles are homogenously dispersed on the  $MnO_2$  PHMs. In contrast, the great rough dots of agglomerated Ag clusters appeared on the surface of PD-0.3%Ag/MnO<sub>2</sub> PHMs are

evidenced by the SEM and the mapping images (Fig. S1). This further confirms the redox-precipitation method is more efficient to achieve high dispersion of Ag atoms on  $\rm MnO_2$  than photo-deposition method.

Raman test was utilized to further identify the effect of Ag modification on MnO<sub>2</sub> PHMs. As shown in Fig. 3a, both Ag/MnO<sub>2</sub> and MnO<sub>2</sub> PHMs showed two main Raman adsorption peaks. The strong peak around 305 cm<sup>-1</sup> could be attributed to the stretching vibration  $\nu_3$  (MnO, F<sub>2g</sub> mode) in the basal plane of [MnO<sub>6</sub>] sheets and it was considered as the symmetric stretching mode of oxygen atoms around Mn ions, while the peak around 633 cm<sup>-1</sup> (D mode) was related to oxygen vacancies due to the presence of Mn<sup>3+</sup> in the MnO<sub>2</sub> lattice [48]. The peak fitting of the F<sub>2g</sub> and D modes in the Raman spectra of both samples were conducted to calculate the ratio (I<sub>D</sub>/I<sub>F2g</sub>), which was linked to the

**Table 2** Physical parameters, chemical and surface compositions of MnO<sub>2</sub> and Ag/MnO<sub>2</sub> PHMs.

| Sample                         | Mn <sup>2+</sup> (%) | Mn <sup>3+</sup> (%) | Mn <sup>4+</sup> (%) | O <sub>L</sub> (%) | O <sub>ads</sub> (%) | O <sub>surf</sub> (%) | Ag° (%) | Ag + (%) | Surface element molar ratio |                                  |                     |
|--------------------------------|----------------------|----------------------|----------------------|--------------------|----------------------|-----------------------|---------|----------|-----------------------------|----------------------------------|---------------------|
|                                |                      |                      |                      |                    |                      |                       |         |          | $Mn^{2+} Mn^{3+} / Mn^{4+}$ | O <sub>L</sub> /O <sub>ads</sub> | Ag°/Ag <sup>+</sup> |
| $MnO_2$                        | 6.79                 | 42.13                | 51.09                | 62.21              | 35.23                | 2.16                  |         |          | 0.96                        | 1.78                             |                     |
| 0.3%Ag/MnO <sub>2</sub>        | 9.96                 | 41.68                | 48.36                | 67.94              | 32.06                | 0.00                  | 46.75   | 53.25    | 1.06                        | 2.12                             | 0.88                |
| 0.3%Ag/MnO <sub>2</sub> (used) | 6.66                 | 42.07                | 51.27                | 63.29              | 35.37                | 1.33                  | 39.71   | 60.29    | 0.95                        | 1.76                             | 0.66                |

oxygen defect sites of the catalysts [49]. The  $I_D/I_{F2g}$  value of pure  $MnO_2$  PHMs and 0.3%Ag/MnO<sub>2</sub> PHMs was 1.75% and 4.89%, respectively. This result suggests the Ag doping induced more intrinsic defects sites and oxygen vacancies in the  $MnO_2$  PHMs, consistent with XPS analysis.

H2-TPR measurements were carried out to explore the reducibility of Ag/MnO<sub>2</sub> PHMs, because high reducibility of catalyst could favor the photothermocatalytic reaction [50]. As shown in Fig. 3b, with the increase of reduction temperature, the sample will undergo the successive reduction of surface adsorbed oxygen species, and the process of  $\mathrm{MnO}_2$  $\rightarrow$  Mn<sub>2</sub>O<sub>3</sub>  $\rightarrow$  Mn<sub>3</sub>O<sub>4</sub>  $\rightarrow$  MnO, respectively [51]. In contrast, the Ag/  $MnO_2$  PHMs has stronger low-temperature reducibility than  $MnO_2$ PHMs, as the temperatures of MnO<sub>2</sub> peaks decrease from 200 °C, 307 °C and 492 °C to 114 °C, 154 °C, and 250 °C, respectively. Peer references indicate that activated hydrogen on the Ag surface can easily migrate to the surface of the MnO<sub>2</sub> PHMs and thus facilitate the reduction reaction at low temperatures [52]. Meanwhile, the incorporated Ag also can activate surface O<sub>L</sub> species of MnO<sub>2</sub> PHMs, which are easier to be desorbed and react with H<sub>2</sub> at low temperature [18,20]. Accordingly, the better reducibility and higher oxygen mobility for 0.3% Ag/MnO2 cause more oxygen to be adsorbed and further excited to reactive oxygen species, which would then be involved in the photothermocatalytic reaction of Ag/MnO2 PHMs.

To further explore the optical performance of the catalysts, the UV–vis-NIR diffuse reflectance (DRS) tests were performed (Fig. 3d). For pure  $MnO_2$  PHMs, a light absorption in the visible light region with the absorption edge at about 650 nm corresponds to the wide bandgap of  $MnO_2$ , and a limited light absorption in the range of 800–1600 nm is attributed to the multiple scattering absorption of  $MnO_2$  porous

structure. In contrast, Ag/MnO<sub>2</sub> PHMs displays an enhanced visible light (400 <  $\lambda$  < 800 nm) absorption resulted from the adsorption band of Ag and stronger infrared adsorption (980 <  $\lambda$  < 1600 nm) attributed to the plasmonic scattering effects of Ag nanoparticles [28].

#### 3.2. Solar light-driven photothermocatalytic bacterial inactivation

The photothermocatalytic disinfection for all the as-prepared catalysts was tested under Xenon lamp. As shown in Fig. 4a, MnO2 and 0.3%Ag/MnO2 PHMs exhibit about 0.1-0.5 log of inactivated E. coli cells in the dark, indicating the catalysts have limited cytotoxicity to E. coli. Upon 10 min Xenon lamp irradiation (309.2 mW cm  $^{-2}$ ), a 0.1% Ag/MnO2 PHMs with rare Ag content can realize 3 log of E. coli inactivation, but MnO<sub>2</sub> PHMs exhibited only about 1.12 log cell reduction within same time period, indicative of enhanced catalytic effect from Ag doping. Moreover, the Ag content on MnO2 and preparation method exhibit great influence on the inactivation of cells. The optimal 0.5% Ag/MnO<sub>2</sub> PHMs can totally inactivate 7.11 log of E. coli within 10 min, while the 0.3%Ag/MnO<sub>2</sub> PHMs with lower Ag doping also obtained the similar inactivation kinetics, suggesting the highly utilization of Ag nanoparticles for 0.3%Ag/MnO<sub>2</sub> PHMs. When the loaded Ag increases from 0.1% to 0.3%, the inactivation efficiency can be enhanced significantly, mainly due to the high dispersion and utilization of atomic Ag. However, when the loading amounts of Ag further increased to 0.5%, the atomic Ag nanoparticles may agglomerate into large particles due to its high surface energy [42,43], thus showing limited enhancement in the E. coli inactivation than the 0.3% Ag loaded sample. Especially, although containing the same content of Ag, the 0.3%Ag/

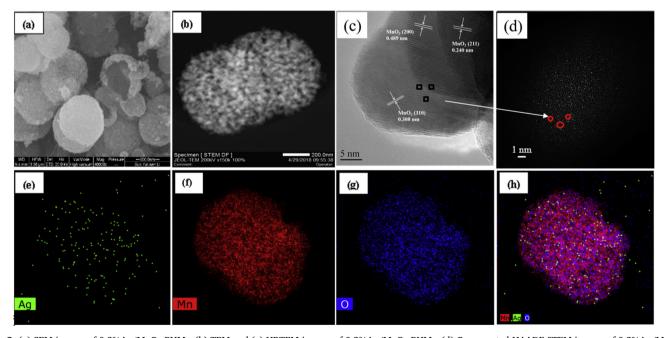


Fig. 2. (a) SEM images of 0.3%Ag/MnO<sub>2</sub> PHMs, (b) TEM and (c) HRTEM images of 0.3%Ag/MnO<sub>2</sub> PHMs, (d) Cs-corrected HAADF-STEM images of 0.3%Ag/MnO<sub>2</sub> PHMs at the same district with inserted partially magnified pictures, (e–h) HAADF-STEM of 0.3%Ag/MnO<sub>2</sub> PHMs with element distribution images corresponding to Ag, Mn, O.

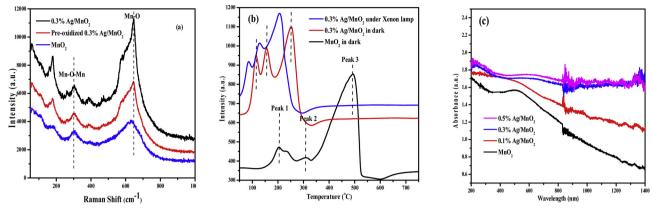
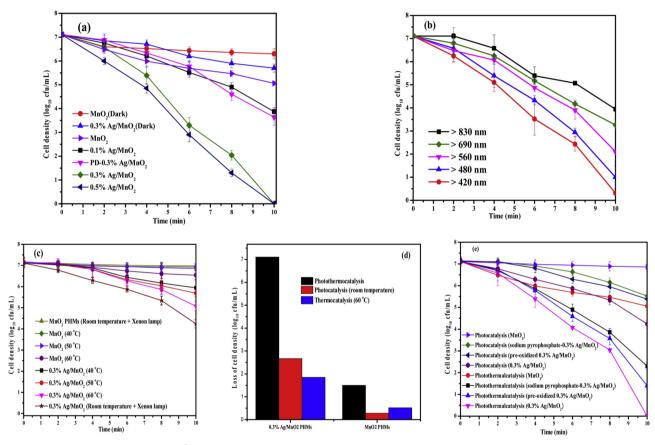


Fig. 3. (a) Raman patterns, (b) H<sub>2</sub>-TPR profiles and (c) UV-vis-NIR adsorption curves of MnO<sub>2</sub> PHMs and 0.3%Ag/MnO<sub>2</sub> PHMs.



**Fig. 4.** Inactivation efficiency against *E. coli* (10<sup>7</sup> cfu/mL) with catalysts, (a) Under Xenon lamp irradiation; (b) with different filter (420, 480, 560, 690 and 830 nm); (c) at room temperature under Xenon lamp, and at different temperature in the dark; (d) Comparison of inactivation efficiency of photothermocatalysis, photocatalysis and thermocatalysis; (e) Inactivation efficiency against *E. coli* by 0.3%Ag/MnO<sub>2</sub> PHMs (role of Mn<sup>3+</sup> and oxygen vacancy).

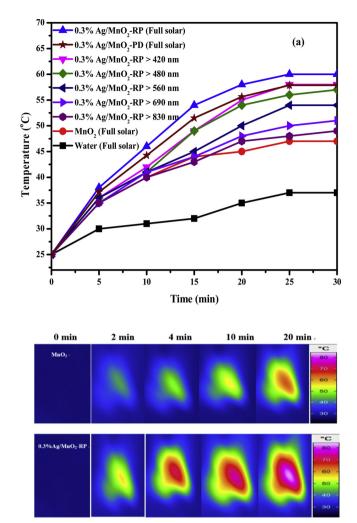
MnO<sub>2</sub> PHMs shows a better activity than PD-0.3%Ag/MnO<sub>2</sub> PHMs prepared *via* photo-deposition (PD), demonstrating the feasibility of redox precipitation to prepare efficient catalyst for water disinfection.

The reaction proceeds through visible-infrared or infrared light with  $0.3\% Ag/MnO_2$  PHMs was also performed under the irradiation of Xenon lamp by using different long wave pass cutoff filters. As shown in Fig. 4b, under the irradiation of visible-infrared light with the wavelength above 420, 480, 560, and 690 nm (the corresponding light intensity is 291.9, 273.3, 255.0, and 238.7 mW/cm², respectively), the corresponding survived cell density of  $0.3\% Ag/MnO_2$  PHMs is 0.5, 1.1, 2.1, and 3.26 log, respectively. Even under the infrared irradiation above 830 nm with a light intensity of  $206.2 \, \text{mW/cm}^2, \, 0.3\% Ag/MnO_2$  PHMs still achieves bacterial inactivation with a residual cell of 3.94

 $\log_{10}$  cfu/mL. These results suggest that  $0.3\% Ag/MnO_2$  PHMs demonstrates UV–vis-NIR, vis-NIR, and NIR catalytic activity with high efficiency.

Due to the strong absorption of Ag/MnO<sub>2</sub> PHMs in the full solar spectrum region (Fig. 3c), there are two possible mechanisms that may contribute to its highly efficient solar light driven *E. coli* inactivation, (1) photocatalysis: electron and hole can be separated by the absorption of photons with energy higher than the band gap of the semiconductor, inducing the redox reaction of the reactive species [52]; (2) thermocatalysis: the surface temperature of the catalyst can be enhanced by the absorption of photons due to photothermal conversion, thus stimulating the thermocatalysis and may also trigger surface reactive species [53]. To clarify this issue, the photocatalytic inactivation of

3%Ag/MnO--PI



**Fig. 5.** (a) Temperature *versus* time plots recorded in water,  $MnO_2$  and  $0.3\%Ag/MnO_2$  PHMs suspension upon irradiation by Xenon lamp (0.25 W cm $^{-2}$ ) with/without different filter (420 nm, 480 nm, 560 nm, 690 nm and 830 nm); and (b) Thermo images for  $MnO_2$  and  $0.3\%Ag/MnO_2$  PHMs.

0.3%Ag/MnO $_2$  PHMs with full solar spectrum irradiation at room temperature was conducted. In this case, about 3.42  $\log_{10}$  cfu/mL cell inactivation was detected (Fig. 4c), indicating that 0.3%Ag/MnO $_2$  PHMs has some photocatalytic activity based on the conventional photocatalysis mechanism. However, no detectable *E. coli* loss was obtained for MnO $_2$  PHMs at room temperature under the irradiation by the Xenon lamp, suggesting that limited photocatalytic activity of MnO $_2$  PHMs. Obviously, the deposited atomic Ag even with 0.1% can efficiently enhance the photocatalytic inactivation of *E. coli*, mainly caused by the narrowed bandgap of MnO $_2$  after Ag doping. This means that the highly efficient bacterial inactivation may also originate from the solar light-driven thermocatalysis, *i.e.* Ag/MnO $_2$  absorb the solar energy, and transform the absorbed solar energy to thermal energy, leading to an increase in the temperature and form reactive species of the Ag/MnO $_2$  suspension.

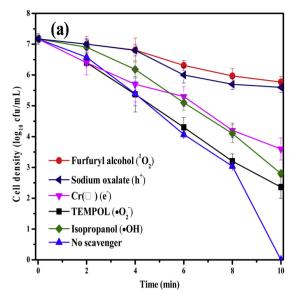
To identify the solar light driven thermocatalysis, the photothermal conversion ability of the catalysts was studied firstly. The solution

temperature evolution of the catalysts with the irradiation of Xenon lamp under the conditions identical to the photocatalytic test was measured. As shown in Fig. 5a, the full solar spectrum irradiation leads to a rapid increase in the temperature of MnO2 PHMs solution to an equilibrium temperature (Teq) of 46 °C at which the equilibrium between the absorption of photon energy by MnO2 PHMs and the dissipation of energy to the surroundings is established. The temperature increase upon solar-light irradiation can thus be attributed to the heating effect of the energy released by the quick non-radiative recombination of the electron-hole pairs produced by the d-d transitions of Mn ions in MnO<sub>2</sub> PHMs upon the absorption of photons [54]. In contrast, the  $T_{\rm eq}$  of 0.3% Ag/MnO<sub>2</sub> PHMs (61 °C) is much higher than that of MnO<sub>2</sub> PHMs, indicating that the strong surface plasmonic absorption of Ag nanoparticles in Ag/MnO2 PHMs makes a considerable contribution to the surface temperature increase. Besides the local heating effect of atomic Ag, its excellent thermal conductivity and low heat capacity also can accelerate the rise of temperature through the heat diffusion from  $MnO_2$  to Ag [18]. Moreover, the  $T_{eq}$  of 0.3%Ag/ MnO<sub>2</sub> with visible-infrared or infrared irradiation above 420, 480, 560, 690, and 830 nm were also measured; the corresponding T<sub>eq</sub> is 58, 57, 54, 51, and 49 °C, respectively (Fig. 5a). It should be noted that pure water displays a limited enhancement in temperature (35 °C) under the same irradiation conditions, which is attributed to the heating effect from the infrared irradiation with the Xenon lamp. Therefore, the results indicate atomic Ag doping can enhance the photothermal conversion of MnO2, and it can proceed with full solar, visible-infrared and even infrared light. Moreover, as shown in Fig. 5a, the Tea of both 0.3%Ag/MnO<sub>2</sub>-PR (61 °C) is much higher than that of 0.3%Ag/MnO<sub>2</sub>-PD (50 °C) MnO<sub>2</sub> PHMs, indicating the high dispersion of atomic Ag is beneficial for the temperature increasing. Furthermore, the further experiments of individual Ag catalysis were carried out with the irradiation of Xenon lamp under the same conditions, the solution temperature evolution was tested, and the result was showed in Fig. S4. Compared with water solution only (37 °C), the solution temperature was merely promoted with 0.03 mg Ag (37.5 °C) and 3 mg Ag (41 °C). Therefore, the individual Ag also has some contribution to the whole photothermocatalytic reaction.

Additionally, to directly investigate the surface photothermal effect for 0.3%Ag/MnO<sub>2</sub> PHMs in converting light into heat, the IR thermocamera was also utilized. As shown in Fig. 5b, the color of images was varied with the degree of temperature and the right scale is corresponding to different color. As shown in Fig. 5b, With the increase of irradiation time, the color of 0.3%Ag/MnO<sub>2</sub> PHMs-RP images became brighter and temperature rose significantly to over 80 °C, while lower temperature increased for the 0.3%Ag/MnO<sub>2</sub> PHMs-PD (near 80 °C) and the MnO<sub>2</sub> PHMs (70 °C), further confirming the increment of thermal energy was mainly attributed to its highly dispersed atomic Ag rather the elemental Ag cluster.

Furthermore, to study the contribution of thermocatalysis for E. coli inactivation, the inactivation efficiency of 0.3% Ag/MnO2 PHMs suspension at the different controlled temperature in dark was also measured. As shown in Fig. 4c, the thermocatalysis activity of pure MnO<sub>2</sub> PHMs is very low, as no significant cells' loss can be observed even increasing the solution temperature from 40 to 60 °C within 10 min reaction. In contrast, loading Ag on MnO2 leads to a considerable improvement in thermocatalytic activity. When the reaction temperature increases above 40 °C, E. coli starts to be inactivated. Further increase the temperature to 60 °C, almost 1.85 log<sub>10</sub> cfu/mL of cells loss was achieved. This result indicates thermocatalytic E. coli inactivation can proceed for 0.3%Ag/MnO<sub>2</sub> PHMs and atomic Ag doping can enhance the thermocatalytic ability of MnO2 PHMs. The atomic Ag can induce higher reducibility for MnO<sub>2</sub> (Fig. 1b), it results in much higher activity of lattice oxygen in MnO2, thus promoting the thermocatalysis to inactivate E. coli under higher temperature.

Do the photocatalysis and the solar light driven thermocatalysis on the 0.3%Ag/MnO<sub>2</sub> PHMs occur independently for *E. coli* inactivation?



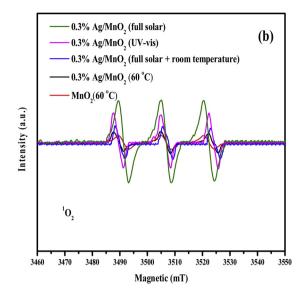


Fig. 6. (a) Inactivation efficiency against *E. coli* ( $10^7$  cfu/mL) by 0.3%Ag/MnO<sub>2</sub> PHMs with different scavengers (0.5 mM Cr(VI), 1 mM isopropanol, 1 mM TEMPOL, 1 mM FFA, and 1 mM sodium oxalate, respectively); (b) ESR spectra of  $^1$ O<sub>2</sub> generated by 0.3%Ag/MnO<sub>2</sub> PHMs under full solar spectrum and UV–vis irradiation, as well as under dark condition, respectively.

To clarify this issue, the comparison in the *E. coli* inactivation efficiency was depicted in Fig. 4d. It can be found that the *E. coli* inactivation efficiency under full solar light (7.11  $\log_{10}$  cfu/mL of cell loss) is much higher than the corresponding summation of the photocatalysis (2.67  $\log_{10}$  cfu/mL of cell loss, room temperature) and the thermocatalysis even at the reaction temperature of 60 °C in dark (1.85  $\log_{10}$  cfu/mL of cells loss). This result clearly indicates the existence of a synergetic effect of the photocatalysis and thermocatalysis on the 0.3%Ag/MnO<sub>2</sub> PHMs under the full solar spectrum irradiation. Similarly, the synergistic effect of thermocatalysis (0.52  $\log_{10}$  cfu/mL) and photocatalysis (0.28  $\log_{10}$  cfu/mL) was also observed for MnO<sub>2</sub> PHMs with 1.5  $\log_{10}$  cfu/mL cells' loss under solar light irradiation. Obviously, the much higher photocatalytic activity and more efficient thermocatalysis of 0.3%Ag/MnO<sub>2</sub> PHMs results in its much higher photothermocatalytic activity than MnO<sub>2</sub> PHMs under the full solar irradiation

Because  $MnO_2$  only exhibits limited photocatalytic bactericidal activity (0.52  $\log_{10}$  cfu/mL for photothermalcatalysis; 0.28  $\log_{10}$  cfu/mL for photocatalysis), the contribution from modified Ag,  $Mn^{3+}$ , oxygen vacancies or the silver itself were further analyzed. In contrast, through using the performance of 0.3%Ag/MnO<sub>2</sub> PHMs to subtract the performance of MnO<sub>2</sub> PHMs, the deduced photothermalcatalytic and photocatalytic inactivation efficiency by the modified 0.3%Ag is 1.85  $\log_{10}$  cfu/mL and 2.39  $\log_{10}$  cfu/mL (Fig. 4d).

About the role of  $\rm Mn^{3+}$ , tetrasodium pyrophosphate ( $\rm Na_4P_2O_7$ ), a widely-used scavenger was added to complex  $\rm Mn^{3+}$  [55–57]. As shown in Fig. 4e, the inactivation efficiency of  $\rm 0.3\% Ag/MnO_2$  is inhibited to some content (4.8  $\rm log_{10}$  cfu/mL for photothermalcatalysis; 0.6  $\rm log_{10}$  cfu/mL for photocatalysis). The deduced photothermalcatalytic and photocatalytic inactivation of *E. coli* by  $\rm Mn^{3+}$  is 2.3  $\rm log_{10}$  cfu/mL and 1.085  $\rm log_{10}$  cfu/mL, which clearly confirms the photothermalcatalytic and photocatalytic role of  $\rm Mn^{3+}$ .

About the role of oxygen vacancies, the pre-oxidized  $0.3\% Ag/MnO_2$  sample was used  $(0.3\% Ag/MnO_2$  was previously oxidized by  $O_3$  for 30 min, thus the oxygen vacancy was replenished by O element). References have been reported that the oxidation reaction with  $O_3$  can fill up the oxygen vacancies [58,59]. Moreover, the Raman results (Fig. 3a) also confirmed lower oxygen vacancies in the pre-oxidized  $0.3\% Ag/MnO_2$  PHMs, since the  $I_D/I_{F2g}$  value (3.006%) of pre-oxidized  $0.3\% Ag/MnO_2$  PHMs was lower than that of fresh  $0.3\% Ag/MnO_2$  PHMs (4.89%), respectively. As shown in Fig. 4e, the inactivation efficiency of

pre-oxidized  $0.3\% Ag/MnO_2$  PHMs (5.71  $log_{10}$  cfu/mL for photothermalcatalysis; 1.73  $log_{10}$  cfu/mL for photocatalysis) was greatly inhibited. The result clearly confirmed the photothermalcatalytic and photocatalytic bactericidal role of oxygen vacancy, as the deduced photothermalcatalytic and photocatalytic inactivation of *E. coli* by oxygen vacancy is 1.4  $log_{10}$  cfu/mL and 0.94  $log_{10}$  cfu/mL.

In comparision, the photothermal catalytic and photocatalytic bactericidal role is in the rank of modified atomic Ag (5.61  $\log_{10}$  cfu/mL, 2.39  $\log_{10}$  cfu/mL) >  ${\rm Mn^{3}}^+$  (2.3  $\log_{10}$  cfu/mL, 1.085  $\log_{10}$  cfu/mL) > oxygen vacancy (1.4  $\log_{10}$  cfu/mL, 0.94  $\log_{10}$  cfu/mL) > MnO<sub>2</sub> (1.5  $\log_{10}$  cfu/mL, 0.28  $\log_{10}$  cfu/mL) > elemental Ag (0.34  $\log_{10}$  cfu/mL). The results clearly confirmed the critical role of atomic Ag to the enhanced photothermal catalytic and photocatalytic activity of MnO<sub>2</sub> PHMs

To further investigate the inactivation capability of  $0.3\% Ag/MnO_2$  PHMs under simulated authentic water conditions, different initial pH and cell density experiments were also performed. As shown in Fig. S2a,  $0.3\% Ag/MnO_2$  PHMs exhibited wide-pH suitability and only slightly higher efficiency was observed at pH 9.29 than 5.60, 7.32. The better inactivation under alkaline condition is mainly due to the more easy generation of reactive species like 'OH generation [7]. As depicted in Fig. S2b, with the increase of initial cell density from  $3\log_{10}$  cfu/mL to  $9\log_{10}$  cfu/mL,  $0.3\% Ag/MnO_2$  PHMs also can maintain good inactivation capability with the prolonged irradiation time from 2 min to 14 min.

When illuminated with solar light, the photothermal conversion of  $Ag/MnO_2$  PHMs gives rise to the rapid increase in the temperature of supported Ag, which may trigger the fast release of  $Ag^+$  [18]. To identify this, the released amount of  $Ag^+$  was measured (Fig. S3). However, only limited  $Ag^+$  (ca. 4.2 ppb) can be determined and no cell loss with the addition of  $5\,\mu g/L$  of  $Ag^+$  in the control experiment, indicating the antibacterial activity was attributed to the photothermocatalytic effect of  $0.3\% Ag/MnO_2$  rather than the released  $Ag^+$ . In contrast, over 26 ppb  $\mu g/L$  of  $Ag^+$  ions were released by  $0.3\% Ag/MnO_2$ -PD. This also confirms the good stability of atomic  $Ag/MnO_2$ -PHMs prepared by redox-preparation.

#### 3.3. Role of various reactive species

The efficient photothermal conversion generated heat by  $Ag/MnO_2$  PHMs can trigger the thermocatalysis to inactivate *E. coli*; it may also

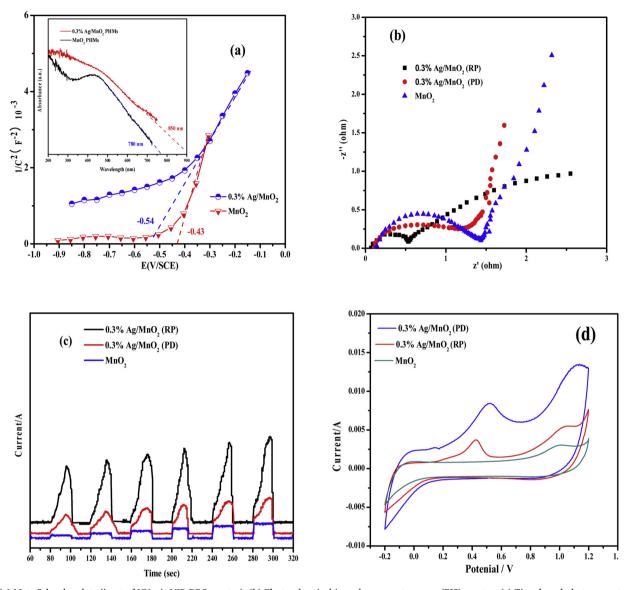


Fig. 7. (a) Mott-Schottky plots (inset of UV-vis-NIR DRS spectra), (b) Electrochemical impedance spectroscopy (EIS) spectra, (c) Time-based photocurrent response and (d) Cyclic voltammetry (CV) curves of MnO<sub>2</sub> and 0.3%Ag/MnO<sub>2</sub> PHMs.

enhance the photocatalysis to form more free radicals to attack E. coli. Trapping experiments were first used to evaluate the role of generated reactive species during the reaction by adding specific chemical scavengers, including Cr(VI) (e<sup>-</sup>), sodium oxalate (h<sup>+</sup>), isopropanol ('OH), furfuryl alcohol ( $^{1}O_{2}$ ) and TEMPOL ( $^{\cdot}O_{2}^{-}$ ) [7,8,14]. As shown in Fig. 6a, compared with that of no scavenger, the E. coli inactivation efficiency of 0.3% Ag/MnO2 PHMs exhibited obviously decrease after the addition of sodium oxalate, indicating the major bactericidal role of h<sup>+</sup>. In contrast, the addition of Cr(VI) to trap e<sup>-</sup> also shows some effect on the disinfection, suggesting the moderate bactericidal role of e-. Besides, some inhibited antibacterial activity also occurs in the presence of TEMPOL and isopropanol, suggesting the limited antibacterial role of  ${}^{\cdot}O_2^{\phantom{*}}$  and  ${}^{\cdot}OH$  species. Actually, based on the Schottky curve in Fig. 7a, it is reasonable to propose that the electrons in the CB of 0.3% Ag/MnO2 PHMs can reduce the absorbed oxygen to generate 'O2"  $(E_0(O_2/O_2^-) = -0.33 \text{ eV } \text{vs NHE})$  and  $H_2O_2$ , which subsequently undergo facile disproportionation to produce 'OH [7,8]. Moreover, plasmonic electrons generated by exciting the Ag NPs also can inject into the CB of MnO2, reducing the surface oxygen to forming 'O2- and OH radicals *via* chain reactions [14]. Importantly, the inactivation of *E*. coli was almost totally inhibited by adding furfuryl alcohol (<sup>1</sup>O<sub>2</sub>, FFA,

 $k = 1.2 \times 10^8 \,\mathrm{M}^{-1} \,\mathrm{s}^{-1}$ ), suggesting  $^{1}\mathrm{O}_{2}$  was the critical reactive species in the reaction [7]. As shown in Fig. 6b, it is worth noting that the signal of DMPO degradation with 1:1:1 characteristic intensity has been observed for the 0.3% Ag/MnO<sub>2</sub> PHMs under Xenon lamp irradiation, which is ascribed to the production of  ${}^{1}O_{2}$  [7]. Especially, the intensity of <sup>1</sup>O<sub>2</sub> signal from full solar light was stronger than that of room temperature and UV-vis alone, indicative of the enhanced generation of reactive species by the photothermal effect. Notably, under dark condition, both 0.3% Ag/MnO<sub>2</sub> PHMs and MnO<sub>2</sub> PHMs can generate <sup>1</sup>O<sub>2</sub> at 60 °C, suggesting the heat can trigger themocatalysis to generate reactive oxygen species for the observed cell inactivation in Fig. 4c. The much higher intensity of  $^{1}O_{2}$  under full light irradiation than that of room temperature with full light irradiation or dark condition, further confirming the synergism of photocatalysis and thermocatalysis in 0.3%Ag/MnO<sub>2</sub> PHMs. As identified above, <sup>1</sup>O<sub>2</sub> and h<sup>+</sup> were the major reactive species and responsible for the inactivation of E. coli.

#### 3.4. Mechanism of enhanced solar light driven E. coli inactivation

Based on the considerable enhancement of *E. coli* inactivation performance by Ag/MnO<sub>2</sub> PHMs, it is reasonable to conclude the critical

role of atomic Ag. The Mott-Schottky (MS) plots were first utilized to analyze the effect of single-atom Ag doping to MnO2. As shown in Fig. 7a, the flat band potential of MnO2 and 0.3%Ag/MnO2 PHMs is  $-0.54\,\mathrm{V}$  and  $-0.43\,\mathrm{V}$ , respectively, which is close to the conduction band (CB) edge [14]. Based on the calculated band gap (1.55 and 1.46 V, inset of Fig. 7a), the valence band (VB) of MnO<sub>2</sub> and 0.3%Ag/ MnO<sub>2</sub> PHMs could be estimated to be 1.01 and 1.03 V, respectively. This result indicated that atomic Ag doping can narrow the band gap of MnO<sub>2</sub>, beneficial for the electron-hole separation in 0.3%Ag/MnO<sub>2</sub> PHMs. Moreover, electrochemical tests were applied to further identify the accelerated migration and interface reaction ability of the charges in catalysts [60.61]. Generally, a smaller arc size in electrochemical impedance spectra (EIS) reflects lower charge transfer resistance on the electrode surface [62]. As shown in Fig. 7b, a smaller arc radius was observed for 0.3%Ag/MnO<sub>2</sub> PHMs than that of MnO<sub>2</sub> PHMs, indicating the separation and transfer efficiency of photogenerated e-h+ was greatly enhanced through the Ag-MnO2 interface. In addition, the photocurrent response of the 0.3%Ag/MnO2 PHMs was also significantly increased and maintained with several on-off cycles in comparison to MnO<sub>2</sub> PHMs (Fig. 7c), further confirming that the atomic Ag doping owns a fast and stable interfacial charge transfer [62]. Moreover, as shown in Fig. 7c, photocurrent response of 0.3%Ag/MnO<sub>2</sub> PHMs exhibited slightly temperature-improved within 5 min. This increase in photocurrent is consistent with the increase of the electron lifetime [63], further confirming that the temperature increase accelerates charge transfer.

Similar phenomena was also observed in the CV curves of Fig. 7d, the current of Ag/MnO<sub>2</sub> PHMs are much higher than that of MnO<sub>2</sub> PHMs, verifying the presence of higher surface charge transfer on Ag/MnO<sub>2</sub> PHMs [62]. It should be noted that the contact of atomic Ag nanoparticles on MnO<sub>2</sub> PHMs greatly determine the photothermalcatalytic activity with the reason that the close contact is the key to achieving efficient charge transfer. As identified by Fig. 7b–d, the electron transfer efficiency from MnO<sub>2</sub> to Ag cluster (Ag/MnO<sub>2</sub> PHMs-PD) is much lower in comparison with that from MnO<sub>2</sub> to atomic Ag (Ag/MnO<sub>2</sub> PHMs-RP). It is reasonable to propose that the interfacial contact area in 0.3%Ag/MnO<sub>2</sub> PHMs-PD, which leads to the efficient charge transfer through maximum utilization of Ag NPs due to its higher dispersion of atomic Ag.

Upon solar light irradiation, the hot electrons in  $MnO_2$  with narrowed band gap can be quickly separated and transferred by highly conductive Ag in realizing the spatial separation of hot electrons and holes. Meanwhile, higher concentrations of lower valent Mn and oxygen vacancies in 0.3% Ag/MnO<sub>2</sub> PHMs (XPS and Raman results in Fig. 3) could also accelerate the separation of electron-hole pairs through providing active sites for surface  $O_L$  to capture electron [63]. Additionally, atomic Ag itself can also generate hot electrons by its plasmonic effect, collectively work with the hot electrons transfer on the Ag-MnO<sub>2</sub> interface to participate in the formation of reactive species and achieve the enhanced photocatalysis for Ag/MnO<sub>2</sub> PHMs. Theoretically, the photogenerated e  $^-$  from CB of 0.3%Ag/MnO<sub>2</sub> PHMs is negative enough to reduce  $O_2/H_2O$  into  $^1O_2$  (-0.11 V, [7]), resulting in the critical role of  $^1O_2$  in Fig. 6a. However, the photogenerated h  $^+$ 

from VB of 0.3%Ag/MnO<sub>2</sub> PHMs is not positive enough to oxidize O<sub>2</sub>/H<sub>2</sub>O into 'OH (2.8 V, [14]), causing the limited bactericidal role of 'OH. Instead, the h<sup>+</sup> is still powerful to directly attack *E. coli*, consistent with the results in Fig. 6a.

Simultaneously, the solar light-driven thermocatalysis also occurs on the Ag/MnO<sub>2</sub> PHMs. It is well known that the thermocatalytic oxidation on MnO2 proceeds via the Mars-van Krevelen mechanism: molecules adsorbed on the surface of MnO<sub>2</sub> are oxidized by its surface O<sub>L</sub>, and the produced oxygen vacancies are subsequently replenished by gas-phase O<sub>2</sub> [64]. The reducibility of the surface O<sub>L</sub> in MnO<sub>2</sub> thus plays a decisive role in its thermocatalytic activity because the reduction of MnO<sub>2</sub> is more sluggish than the re-oxidation of reduced MnO<sub>2</sub> [65]. H<sub>2</sub>-TPR in Fig. 3c has confirmed the higher reducibility of MnO<sub>2</sub> PHMs after atomic Ag doping, which means that the O<sub>L</sub> of Ag/MnO<sub>2</sub> PHMs is in a higher activity for thermocatalysis. Moreover, the more efficient photothermal conversion for Ag/MnO2 than MnO2 PHMs was identified in Fig. 5a. The reason is that atomic Ag is with broad absorption bands spanning the UV-vis-NIR wavelengths, which can induce multiple scattering events and increase photon absorption probability, resulting in localized intense heating on MnO2. Also, atomic Ag itself with high thermal conductivity and low heat capacity also can accelerate the rise of temperature through the heat diffusion from MnO<sub>2</sub> to Ag. Therefore, the more efficient photothermal conversion of Ag/MnO2 than MnO2 PHMs together with its higher thermocatalytic activity result in its much higher E. coli inactivation efficiency.

To put more insight in the synergetic effect of photothermocatalysis, the effect of the solar light irradiation on the reducibility of 0.3% Ag/ MnO<sub>2</sub> PHMs was investigated by H<sub>2</sub>-TPR in dark and with the irradiation of the Xenon lamp. As shown in Fig. 3b, notably, compared to H<sub>2</sub>-TPR in dark, the full solar spectrum irradiation of Xenon lamp leads to a considerable shift of H<sub>2</sub> consumption peak to lower temperature, indicative of higher activity of O<sub>L</sub> in Ag/MnO<sub>2</sub> PHMs. This may arise from the following reason: the photogenerated holes in MnO<sub>2</sub> upon full solar irradiation leads to the formation of  $O^-$  species from  $O_L$  ( $h^+$  +  $O_L^{2-} = O^-$ ) and activate  $O_L$  simultaneously, and the active  $O^-$  species may react with H2 to cause the H2 consumption peak shift to lower temperature [66]. Moreover, irradiation with the full solar spectrum resulted in a significant increase in the total H2 consumption amount (Table 3), suggesting that more active O<sub>L</sub> was produced upon solar light irradiation. Therefore, the highly efficient photothermocatalysis is considerably promoted by the synergetic effect through O<sub>L</sub> the hot h<sup>+</sup> generated by photocatalysis can promote the activity and amount of O<sub>L</sub>, thus considerably improving the thermocatalysis activity of 0.3%Ag/ MnO<sub>2</sub> PHMs. Consistently, the consumption of O<sub>L</sub> was observed in the used 0.3%Ag/MnO<sub>2</sub> PHMs in Fig. 1c and Table 2.

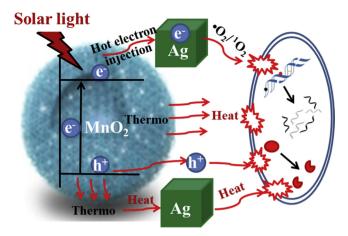
As shown in Scheme 2, with the solar light irradiation of Ag/MnO<sub>2</sub> PHMs, the MnO<sub>2</sub> PHMs not only generate heat through photothermal effect, but also produce hot charge carriers. Both heat and hot electrons then transfer to atomic Ag through Ag-MnO<sub>2</sub> interface, resulting in the elevated temperature as well as separation of electrons and holes to attack cell by heat and reactive species. Meanwhile, the plasmonic Ag itself exhibits strong local heating effect and generate hot electrons in the entire solar irradiation, which also help to kill bacteria by heat and reactive species. Especially, the 3D porous hollow structure of MnO<sub>2</sub>

Table 3
Reduction temperature and H<sub>2</sub> consumption of MnO<sub>2</sub> and Ag/MnO<sub>2</sub> PHMs.

| Catalysts                             | Reduction temperatures (°C) <sup>a</sup> |        |        | ${ m H_2}$ consumptions (mmol/g) $^{ m b}$ |        |        |  |
|---------------------------------------|--|--------|--------|--|--------|--------|--|
|                                       | peak 1                                   | peak 2 | peak 3 | peak 1                                     | peak 2 | peak 3 |  |
| $MnO_2$                               | 200                                      | 307    | 492    | 0.79                                       | 0.68   | 3.34   |  |
| 0.3%Ag/MnO <sub>2</sub>               | 114                                      | 154    | 250    | 1.04                                       | 2.83   | 3.88   |  |
| 0.3%Ag/MnO <sub>2</sub> (irradiation) | 85                                       | 124    | 205    | 1.19                                       | 3.12   | 4.29   |  |

<sup>&</sup>lt;sup>a</sup> Reduction temperatures corresponding to the maximum values of reduction peaks in the H<sub>2</sub>-TPR result.

 $<sup>^{\</sup>mathrm{b}}$  H $_{2}$  consumptions calculated by reduction peak areas.



Scheme 2. Mechanism of E. coli inactivation with  ${\rm Ag/MnO_2}$  PHMs under solar light irradiation.

may provide large active sites and promote the mass transfer process to facilitate the reaction [22,23].

#### 3.5. The bacterial damage process

The morphology of *E. coli* at different stages of the disinfection process was imaged by SEM in Fig. 8a–c. The rod-shaped morphology (Fig. 8a) of *E. coli* cells in the beginning became deformed with some bulges and pits on the surfaces, seriously distorted and fractured with prolong treated (Fig. 8b), indicating that generated reactive species and heat efficiently attack and destroy the cells' envelope. The *E. coli* cells were completely deformed after 10 min treated. Similarly, fluorescent tests were also indicated the damage of *E. coli* cells by 0.3%Ag/MnO<sub>2</sub> PHMs. Green fluorescent stain for both live and dead bacteria, and red fluorescent stain for cells with compromised cellular membranes. The cells red fluorescence increased and nearly completely to replace green

fluorescence in number within 10 min treated, as shown in Fig. 8e and f, indicating that *E. coli* cells were sustainably destructed and debrics were adsorbed on the surfaces of 0.3%Ag/MnO<sub>2</sub> PHMs [7,14].

The oxidative stress mediated by  $0.3\%Ag/MnO_2$  PHMs was examined using glutathione oxidation assay. Glutathione (GSH) is a tripeptide with a thiol group, serves as one of the major cellular antioxidant enzymes in bacteria, with concentration ranging between 0.1 and  $10\,\mathrm{mM}$  [67]. As an indicator of the oxidative stress induced by different nanomaterials, thiol groups (–SH) of glutathione can be oxidized to glutathione (GSH). GSH is involved in the intracellular oxidative balance and protects the cells against external electrophilic compounds, thereby the *E. coli* to quickly synthesize more GSH (from 1.5 to  $2.05\,\mu\mathrm{mmol/L}$ ) to fight against the oxidation and heat attack in the first 5 min treatment (Fig. 9a). However, with prolonged treatment, GSH content in the cells sharply decreased from  $2.05\,\mathrm{to}~0~\mu\mathrm{mmol/L}$  within 30 min treated, indicating the fierce attack from Ag/MnO<sub>2</sub> PHMs exceeded the self-defense capability of *E. coli*.

Even if the cells have been inactivated, some injured cells in the initial phase still can self-repair their respiratory ability and then supply enough energy for regrowth after culture in nutrients [14]. As the key material for storing both vital and direct energy, the synthesis of ATP is directly responsible for cellular metabolic activity [32]. Therefore, ATPase (the ATP synthesis enzyme) was monitored to identify damaged metabolism, whose activity is in proportion to the elevated absorption intensity [64]. As shown in Fig. 9b, accumulated ATP content in the cells decreased with prolong treated (the amount of ATP at an incubation time of 60 min): initial cells contained almost  $1.2\times 10^{-10}$  mmol ATP/cell, which then decreased to  $3\times 10^{-11}$  mmol ATP/cell, and almost no ATP remained after 30 min treated, indicating the cells almost lost all the ability to synthesize ATP after a prolonged treatment period.

Notably, cell still can self-repair and regrow even when their proteins are damaged [64,65]. Only severe damage to the DNA can cause fatal, death of the cells. The leakage and decomposition of genomic DNA could be observed in Fig. 9c–e, which displays that the intensity of

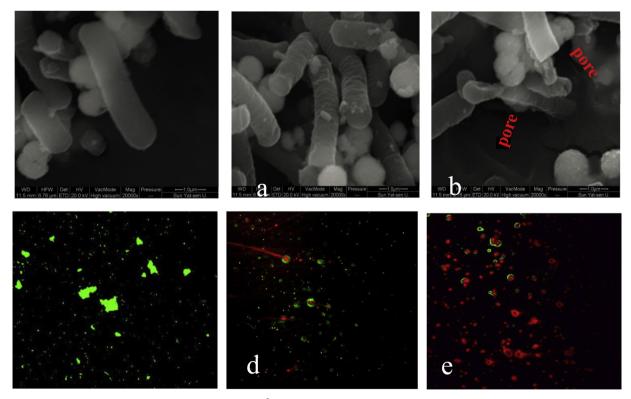
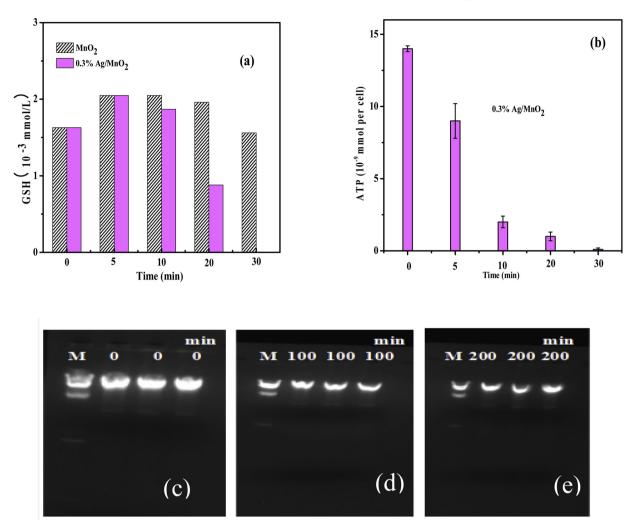


Fig. 8. SEM images and fluorescence microscopic images of *E. coli* (10<sup>8</sup> cfu/mL) treated by 0.3%Ag/MnO<sub>2</sub> PHMs with (a) 0 min, (b) 5 min, (c) 10 min under a Xenon lamp.



**Fig. 9.** The *E. coli* (10<sup>7</sup> cfu/mL) generation potential of GSH (a) and ATP (b) after photothermocatalytic inactivation for 0, 5, 10, 20 and 30 min, and (c–e) the leakage bacterial genomic DNA extracted from harvested cells (10<sup>8</sup> cfu/mL) during photocatalytic inactivation by 0.3%Ag/MnO<sub>2</sub> PHMs with (c) 0 min, (d) 100 min, (e) 200 min.

DNA bands decreased significantly within the 200 min of the process. Within the test time-period, the incomplete elimination of the DNA bands means the total mineralization of *E. coli* by Ag/MnO<sub>2</sub> PHMs has not finished. Although the cell envelope and functional enzymes have been destructed mostly within the test-period, its residual organic fragment will compete with the DNA to consume the ROS and heat. Therefore, it still needs a longer reaction time to totally mineralize the biomolecules and even DNA [16]. In addition, during the bacterial inactivation process, the bacteria division and then the leakage of DNA from the cell were in a dynamic equilibrium, which also hinder the complete destruction of intracellular DNA.

#### 4. Conclusions

In this work, the high dispersion of atomic Ag nanoparticles on  $\rm MnO_2$  porous hollow microspheres (the optimum 0.3%Ag/MnO\_2 PHMs) was prepared by redox-precipitation methods, which achieved highly photothermocatalytic bacterial inactivation under solar light irradiation. Firstly, attributed to the high conductivity of Ag and its induced low valent Mn and oxygen vacancy, the excitation of  $\rm MnO_2$  by solar light produces hot charge carriers and then quickly transfers to Ag nanoparticles through Ag-MnO\_2 interface, transforming into reactive species for photocatalysis. Secondly, the confined atomic Ag exhibits strong local heating effect and induces higher reducibility for MnO\_2, considerably enhances the photothermal conversion and lattice oxygen

activity of MnO<sub>2</sub>, thus promoting the thermocatalysis. Due to the synergistic eff; ect of photocatalysis and thermocatalysis participating in photothermocatalytic reaction, thus the designed 3D Ag/MnO<sub>2</sub> PHMs exhibit superior inactivation efficiency towards *E. coli* under solar light.

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# Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:https://doi.org/10.1016/j.apcatb.2018.12.056.

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